The essential momentum of neutron imaging approaches to reveal water dynamics at the root-soil interface

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Conventional studies of soil water dynamics

Menon et al., Plant & Soil, 2005

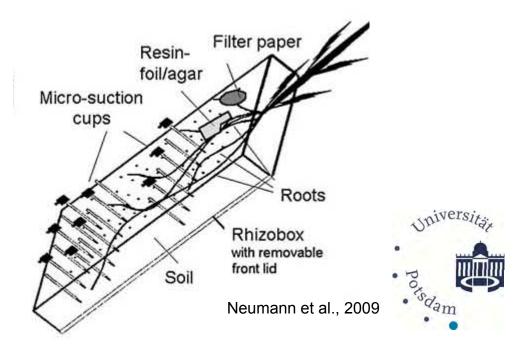


Laboratory columns & lysimeters

Rhizoboxes or the like

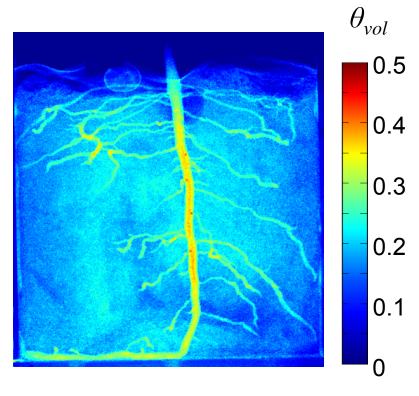


Tensiometer & TDRs



The alternative – Non-invasive Imaging in small rhizobox and column set-ups





- Non-invasive, non-destructive
- Continuous measurements
- Not averaging over a larger local volume



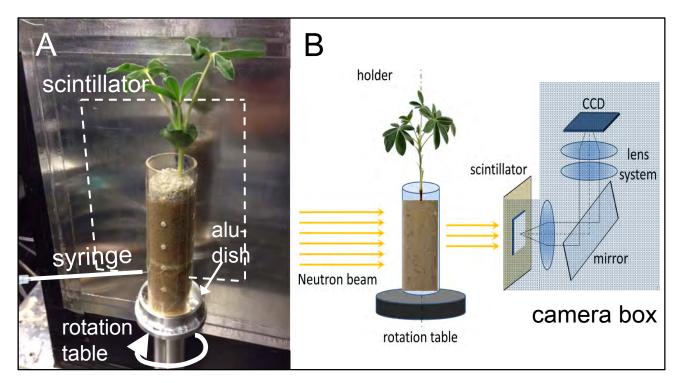
Neutron radiography facilities

Facilities where we have done neutron imaging

- ⇒ Paul Scherrer Institut, Villigen, Switzerland
- ⇒ Helmholtz Centre Berlin, Germany
- ⇒ ILL & Universite Grenoble Alpes, France



www.psi.ch



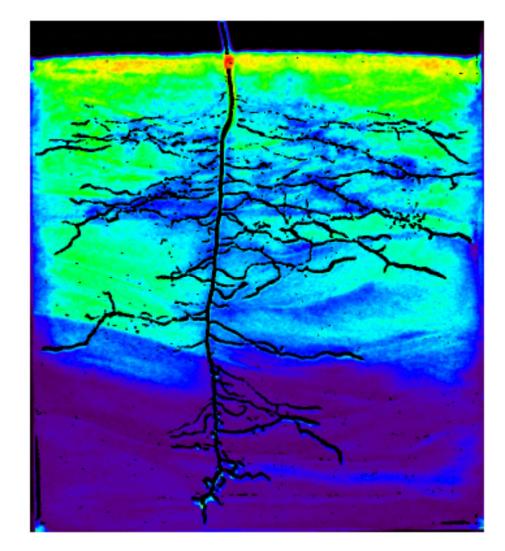


HZB, Berlin

Neutron radiography of a plant-soil system

→ water content distribution and root structure



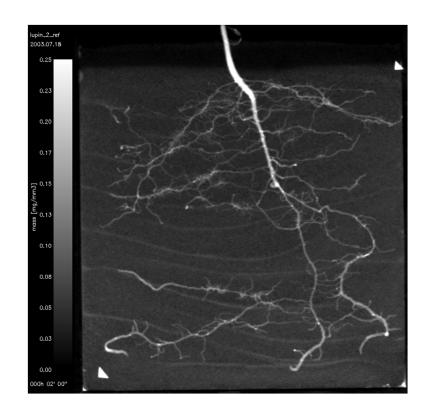


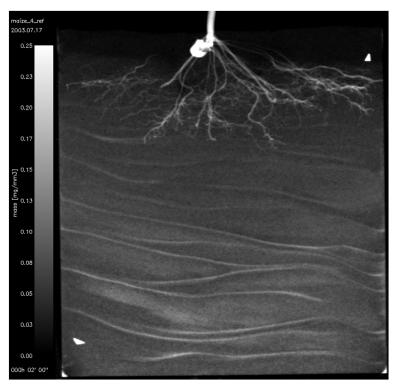
S. Oswald et al., Vadose Zone journal, 2008

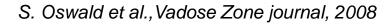


Root growth: lupine

corn

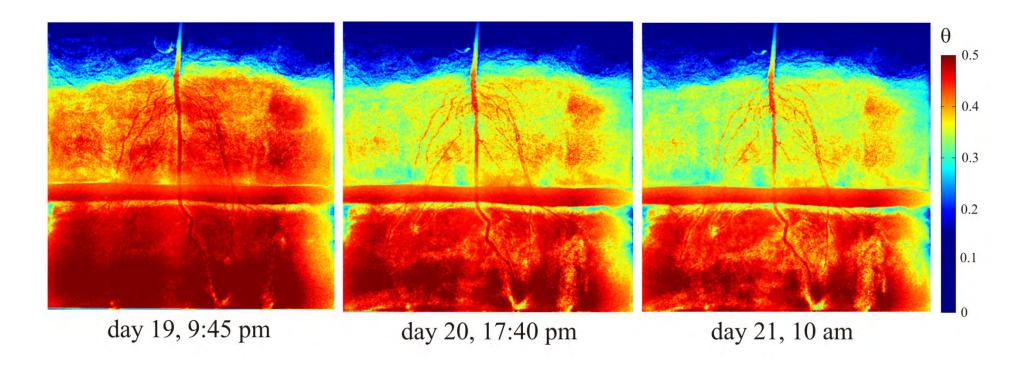








Hydraulic barriers for separation during infiltration and drying





Experiment design for compartments of different water contents

30*25*1 cm³ Aluminum containers



A. Dara et al., Plant & Soil, 2015



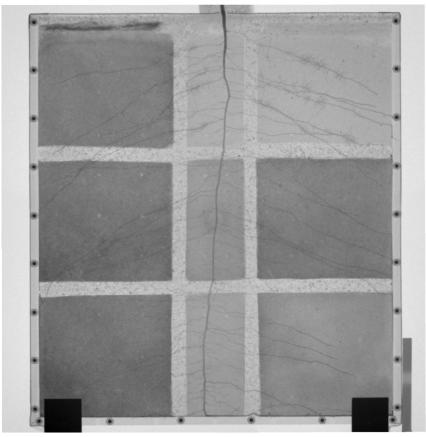
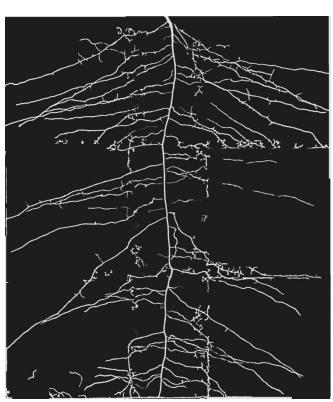


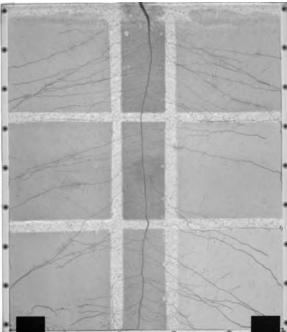
Image analyses to derive local water balance and uptake

A. Dara et al., Plant & Soil, 2015

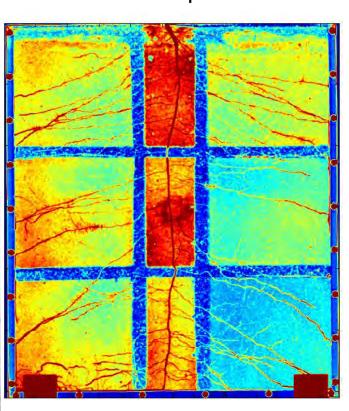
Root segmentation



Raw Radiograph

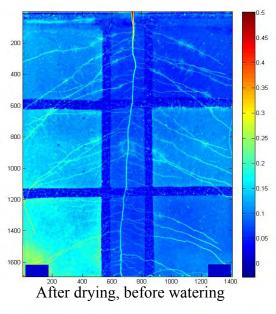


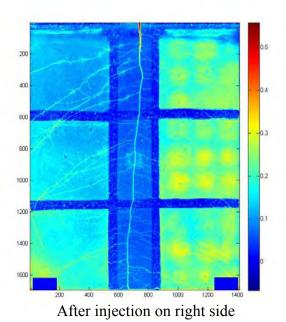
Water content quantification



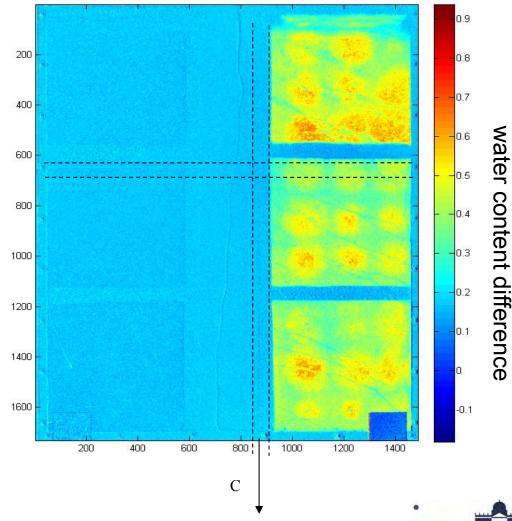


Water balance in individual compartments





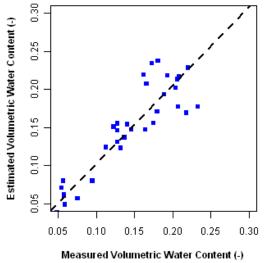
A. Dara et al., Plant & Soil, 2015

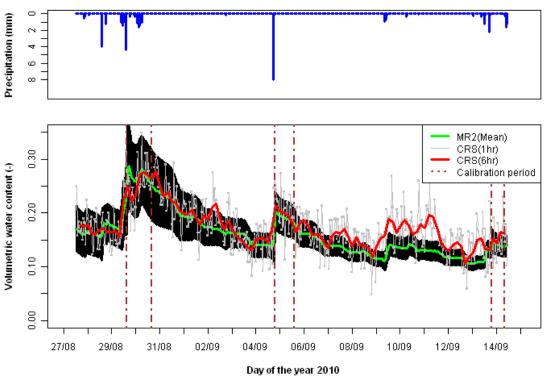


Coarse-sand layer (capillary barrier)

Neutron "imaging" in the field via neutrons produced by cosmic-rays naturally?





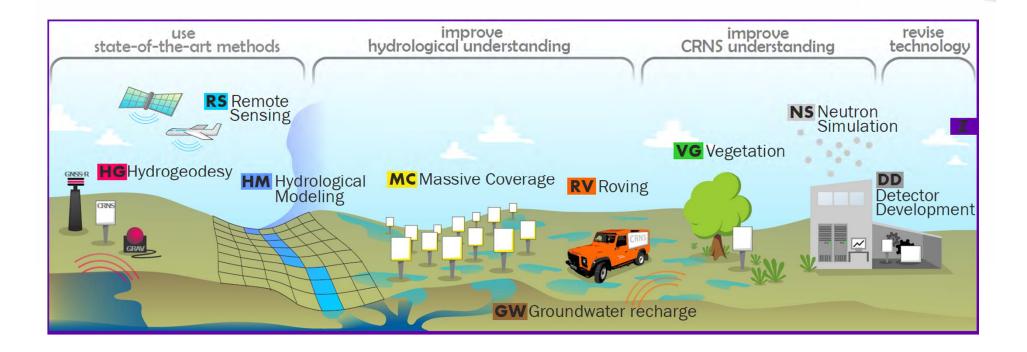


C. Rivera Villarreyes et al., Hydrol. Earth Syst. Sci., 2011



Soil moisture at field scale via neutrons!





Mapping soil moisture via cosmic-ray neutron sensing for the first time - current field monitoring campaign



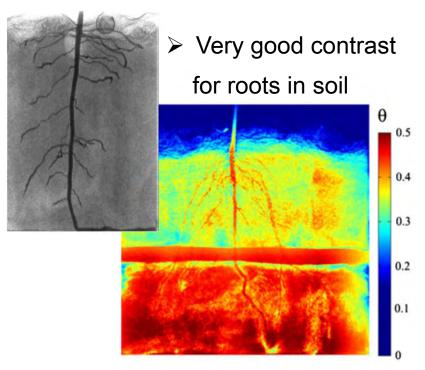
Needs for NI laboratory plant-soil studies

- Controlled environment for growth period and during experiments
- Large(r) samples
- Quicker tomographies with high resolution
- Combination with other imaging methods
 - direct at NI facility
 - same samples
 - similar resolution



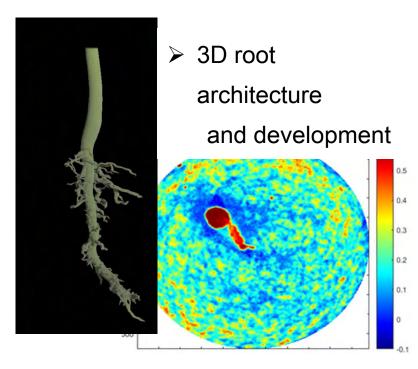
2D vs. 3D imaging mode

Neutron radiography



- > 2D soil water mapping
- Dynamic measurements (tracer injection)

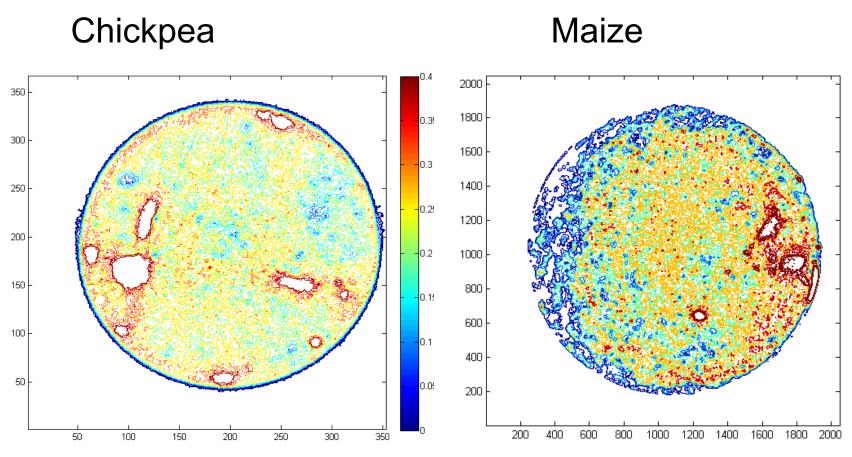
Neutron tomography



- > 3D soil water maps
- No dynamic measurements
 - until recently



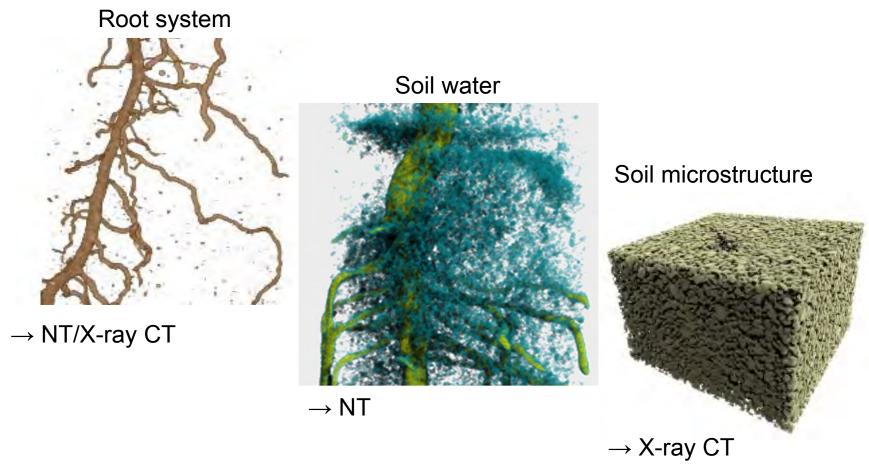
Imaging the rhizosphere behavior in 3D: level 1



Root-microtomography in cylinder by neutron imaging (A. Moradi et al., *New Phytologist*, 2011; showing mucilage effect)



Imaging the rhizosphere behavior in 3D: level 2



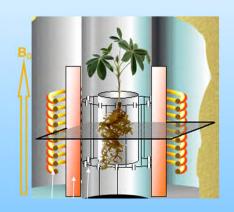
Combining X-ray CT and NT

→ More comprehensive picture of rhizosphere



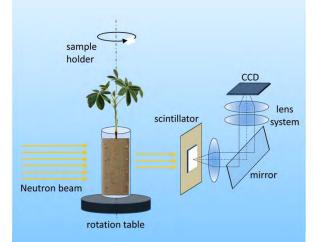
Imaging the rhizosphere behavior in 3D: level 3

Magnetic Resonance Imaging (MRI)



- + voxel signal = f (water, tracer, diffusion, flow, pore structure)
- + thick and wet samples
- relatively insensitive

Neutron Tomography (NT)



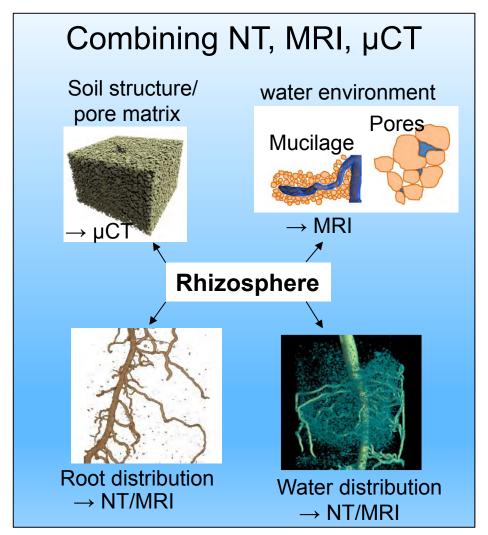
- + high resolution (0.05mm)
- + high sensitivy
- + 3D water distribution
- thin or dry samples

Microfocus X-ray CT (µCT)



- + high resolution (0.05mm)
- + 3D structure of soil matrix
- thin samples
- weak contrast for roots and water

Objectives of combined imaging



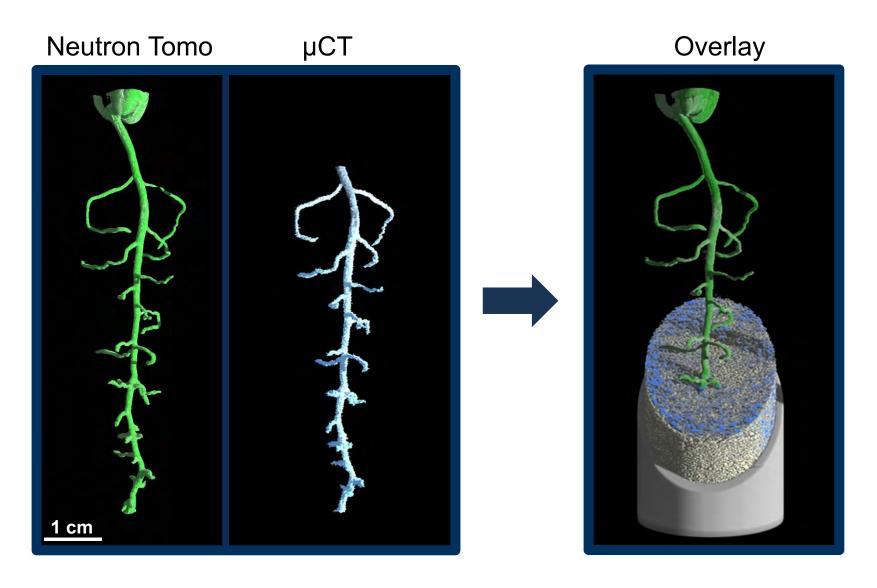


Comprehensive picture of rhizosphere

- Capturing the root system in the same hydro-physiological state
- → novel transportable MRI for direct combination of NI/MRI at the same plant sample
- > Co-registration
- → transformation of different 3D data sets (different resolution, orientation) into a common coordinate system

Christian Tötzke et al., manuscript in preparation

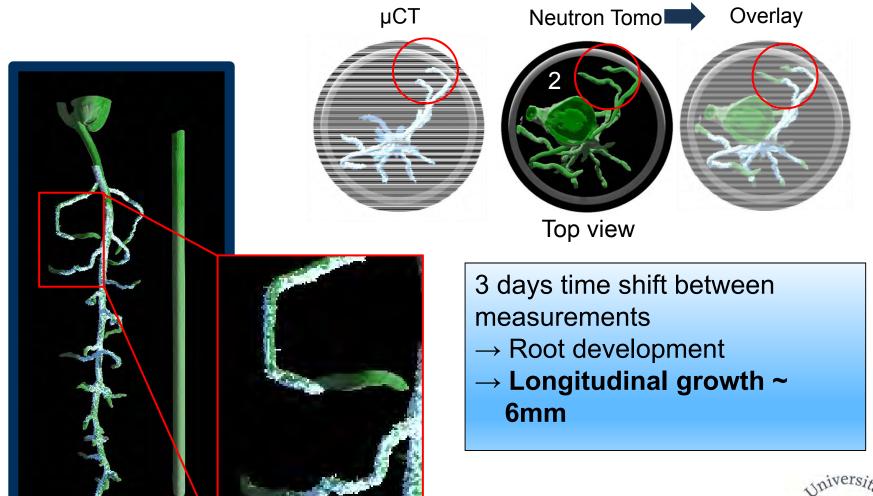
Co-registration – Bean seedling



Christian Tötzke et al., manuscript in preparation

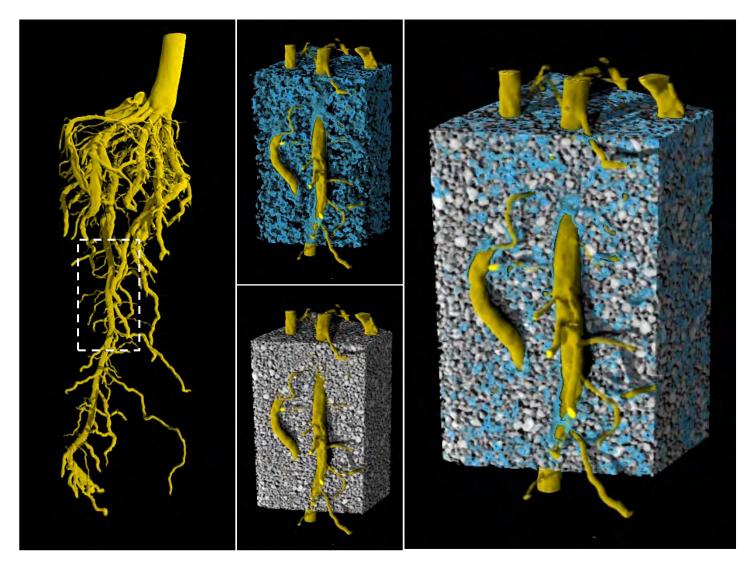
Difference in architecture → Root Growth

1 cm





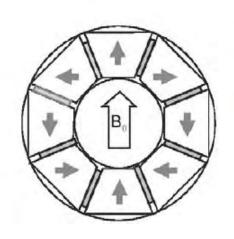
Combined results from Neutrons and X-rays





Christian Tötzke et al., manuscript in preparation, 2019

Mobile MRI possible at NI facility







- Based on permanent magnets
- Very recent development
- MRI sequences and operation by S. Haber-Pohlmeier (RWTH) and A. Pohlmeier (FZJ)



Add MRI relaxation time maps – work together with Sabina Haber-Pohlmeier & Andreas Pohlmeier

Rhizosphere during drying-rewetting cycle

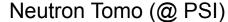
Vicia faba in sand 2 weeks old

Neutron Tomo (@ PSI) Low field MRI (@PSI) θ =0.16cm³/cm θ =0.17cm³/cm θ =0.17cm³/cm After rewetting

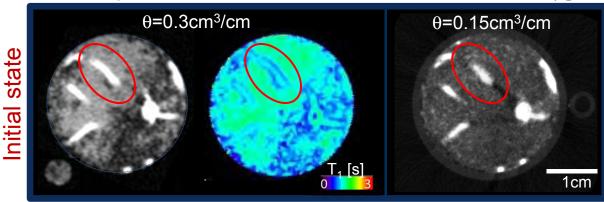
dryer region around root

RARE: rapid acquisition relaxation enhanced H₂O distribution → slow rehydration of the mucilage zone

High field MRI (@ FZ Jülich)



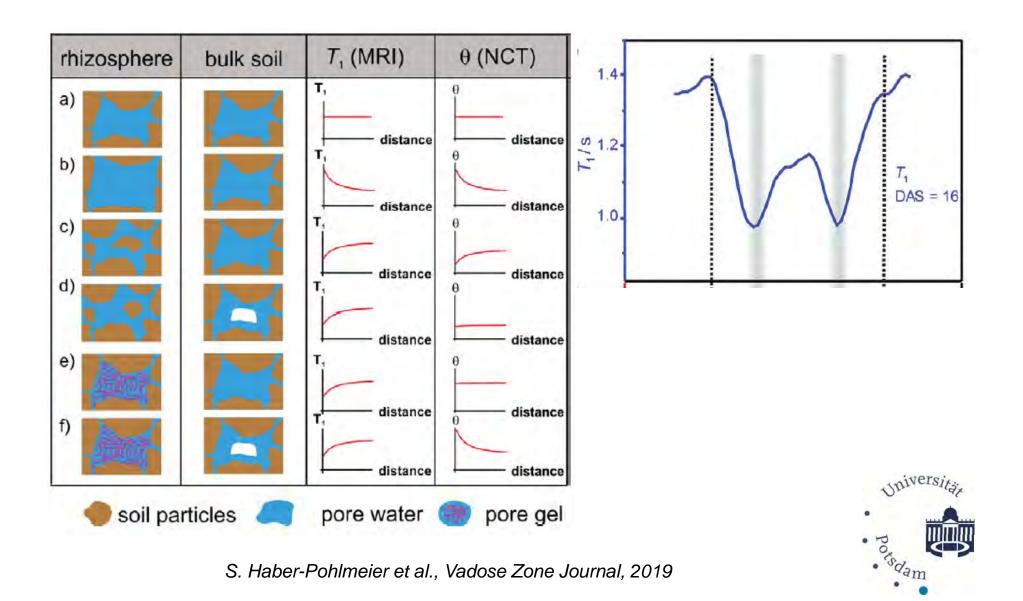
12 h later



T1 map, relaxation time Spin Echo Multi-Slice uniform water distribution →uniform H₂O distribution→ different water environment

Similar in S. Haber-Pohlmeier et al., Vadose Zone Journal, 2019

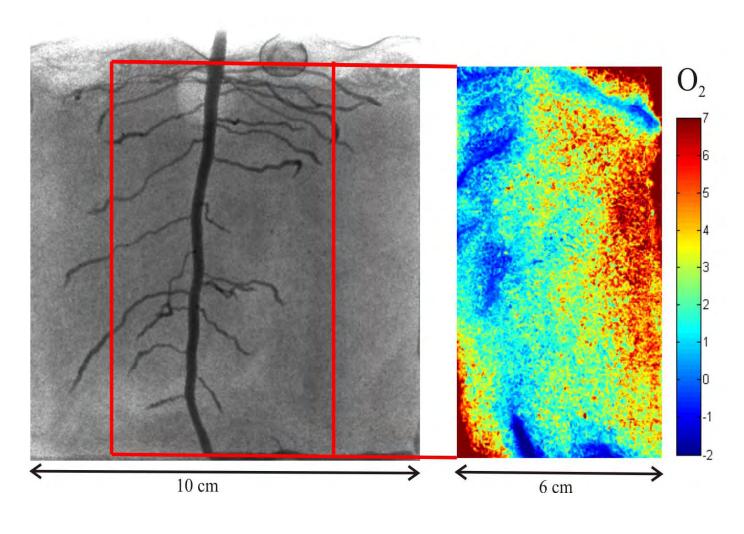
MRI adds relaxation time info (and water)



Combining 2D neutron imaging and Fluorescence Imaging



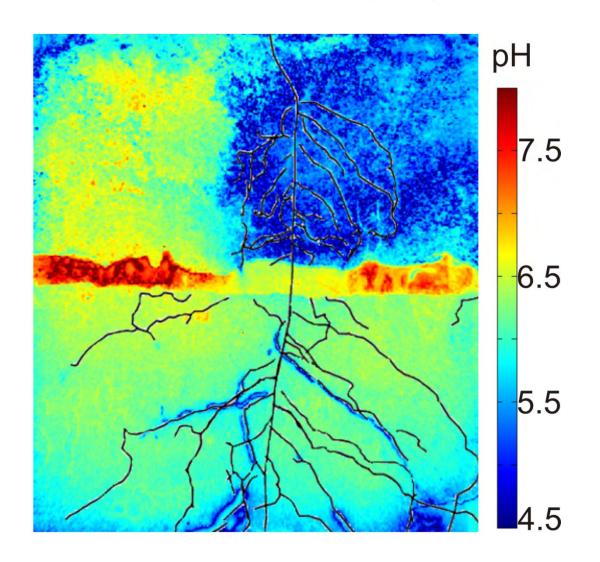
Combined Neutron Imaging and Fluorescence Imaging: Oxygen





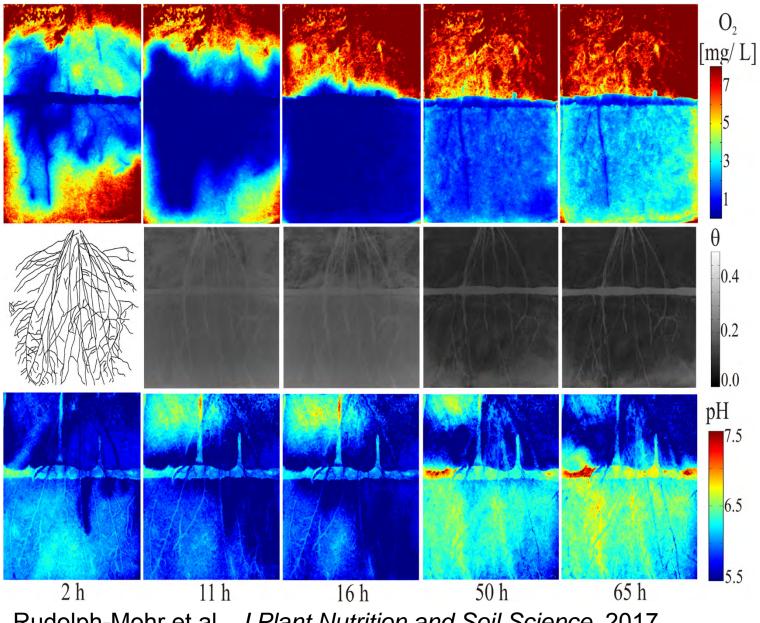
Rudolph et al., J Soils and Sediments, 2012

Combined Neutron Imaging and Fluorescence Imaging: pH





Combined Triple Imaging, water, DO, pH







Summary 2D

- Root growth
- Infiltration and uptake, local rhizosphere effects?
- Water stress compensation in root system
- Combination with fluorescence imaging adds a hydrochemical dimension

but also

- degradation of chemicals in soil aggregates
- Swelling and water storage in soil ammendments

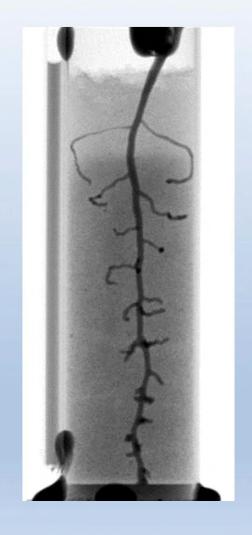


Neutron tomography going dynamic



Capturing 3D water flow in rooted soil

Acquisition time: 100 min



Conventional acquisition > 1 hour

- stepwise sample rotation
- several hundreds radiographic projections
- → quasi-static 3D water distribution
- → too slow to capture 3D water dynamics:
 e.g. water infiltration, root water uptake

Speed!!! < 1 min

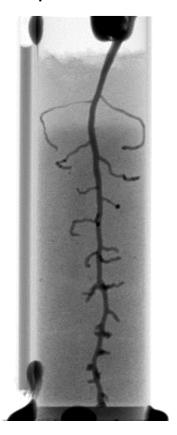
(100 times faster)

Imaging options

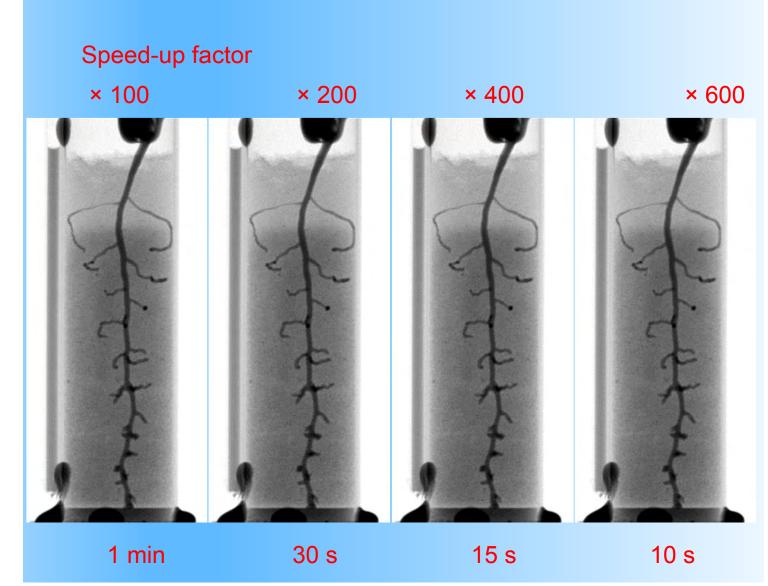
- 1. neutron flux: source, pin hole/distance
- 2. signal per pixel: binning
- 3. rotation: stepwise -> continuous
- 4. number of projections



conventional speed



t = 100 min



Capturing 3D water flow in rooted soil





600 × faster

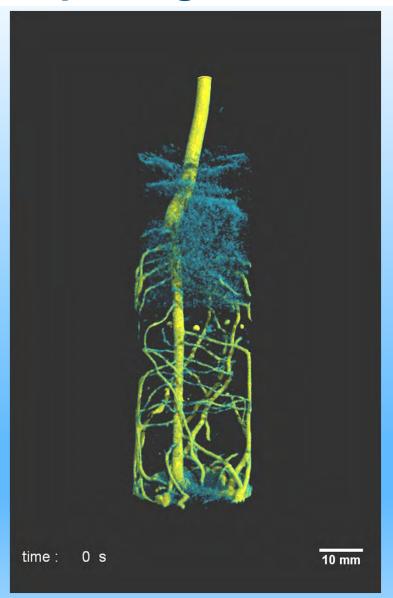
Acquisition time: 10 s

Imaging Parameter

Exposure time/projection = 0.05 s Number of projections = 200 Pixel size = 55 μ m 3x3 Binning \rightarrow 165 μ m CONRAD position: 2 Distance (pinhole-detector) L = 5 m Pinhole D = 3 cm L/D = 167



Capturing 3D water flow in rooted soil - lupin



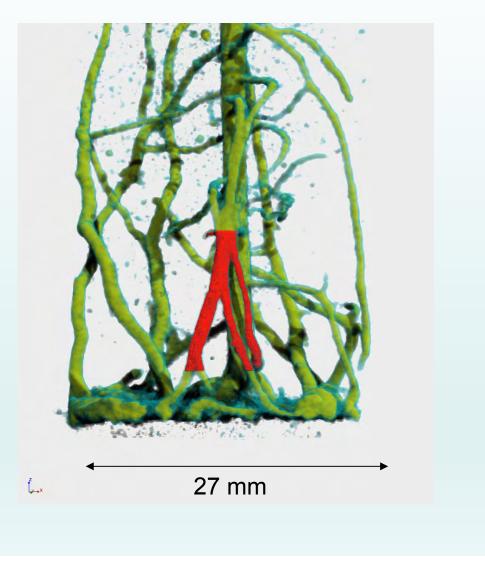
1. D₂O infiltration experiment

Acquisition time: 10 s

- ➤ D₂O injection at bottom
- Replacement of H₂O
- Ascending water front
- Stop at hydraulic barrier



Water (D₂O) uptake by lupine roots



Root uptake



10 min after D₂O injection

Tötzke et al. (2017), Scientific Reports, 7

Water (D₂O) uptake by maize roots



2. D₂O infiltration experiment

Acquisition time: 1 min Resolution: 110 µm/pixel

- > D₂O injection at bottom
- > Ascending water front
- Root uptake

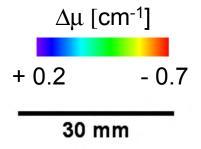


Water (D₂O) uptake by another maize plant



3D maize root system

3. D₂O infiltration experiment + stronger uptake



time 0 min



Summary 3D

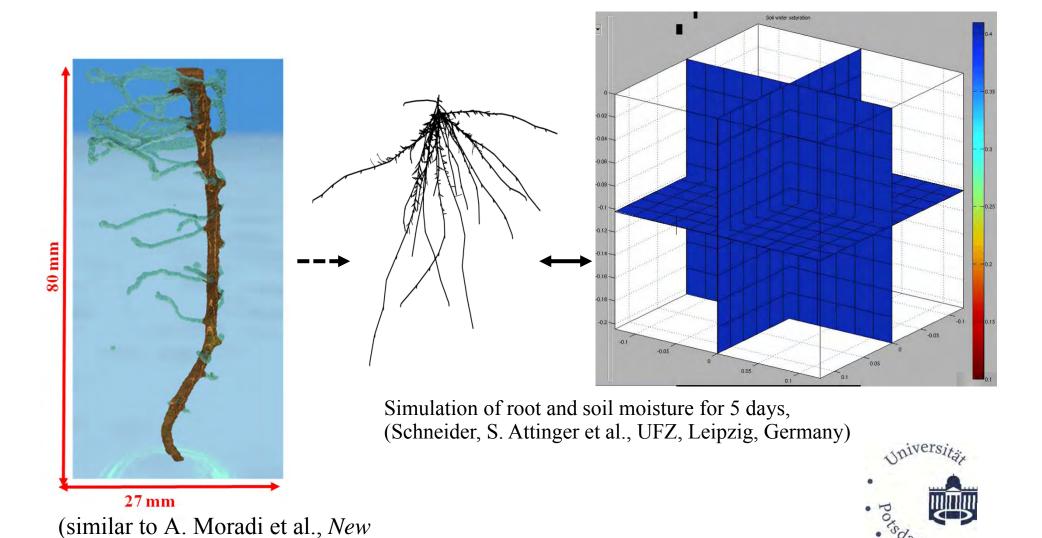
- Combination of NR with XCT and/or MRI possible and beneficial
- ➤ Fast neutron tomography experimental key for studying 3D water dynamics *in situ*
- Infiltration experiments enhance understanding of rhizosphere hydraulics
- Valuable experimental complement for 3D models simulating root water uptake
- Broad applicability of fast NT to analyze 3D transport processes in porous media (materials and geoscience)



Tötzke et al. (2019), in preparation



Missing: combination of root structure and numerical simulation of water flow and root uptake (in 3D)



Phytologist, 2011)

Combining some 2D and 3D advantages: Tomography possible for rectangular container

- Root structure
- Both 2D and 3D
- Water distribution dynamics and local water gradients

